# 2 x Taq Master Mix

Premixed solution for convenient PCR

## **Product Description**

2 x Taq Master Mix is a premixed 2 x concentrated solution of Taq DNA Polymerase (Omega Bio-Tek, Cat. No. TQ2100), Reaction Buffer, MgCl2,dNTPs, stabilizer and enhancer. 2 x Taq Master Mix contains all components for PCR, except DNA template and primers. The mixture is optimized for consistent and efficient routine PCR amplifications. It can amplify up to 8 kb fragment from lambda DNA. For a 50  $\mu$ l reaction, simply add 25  $\mu$ l of 2 x Taq Master Mix to primers, DNA template and PCR-Qualified H2O. In addition, 2 x Taq Master Mix (with dye) can be used to directly load the PCR products onto an agarose gel without the need to add a gel loading buffer.

## **Storage Conditions**

2 x Taq Master Mix should be stored immediately upon receipt at -20°C or 2-8°C. When stored under -20°C and handled correctly, these products can be stored for at least 12 months without showing any reduction in PCR performance. When stored under 2-8°C, 2 x Taq PCR Master Mix Kit can also be stored for up to 2 months.

#### **Kit Components**

	TQ2201	TQ2200
Components	2 x Taq Master Mix	2 x Taq Master
	(with Dye)	Mix (without dye)
Tris	20 mM	20 mM
KCI	100mM	100mM
DNTP	0.4mM	0.4mM
MgCl2	4mM	4mM
Таq	100-200U/ml	50-100U/ml
Dye	Brominecome blue	
Additives	Secret	Secret

#### **Basic PCR Protocol**

This protocol serves as a guideline for PCR amplification. Optimal reaction conditions, such as incubation times, temperatures, and amount of template DNA, may vary and must be individually determined.

- 1. **Thaw primer solutions.** Keep on ice after complete thawing, and mix well before use.
- Mix the Taq Master Mix by vortexing briefly. It is important to mix the Taq PCR Master Mix before use to avoid localized differences in salt concentration.
- 3. Prepare one of the following reaction mixes on ice:
  - For a 25 µl reaction volume:

Component	Volume	Final
Component		Concentration
2X Taq Master Mix	12.5 µl	1X
Upstream Primer, 10 µM	0.5 µl	0.1–1.0 µM
Downstream Primer, 10 µM	0.5 µl	0.1–1.0 µM
DNA Template	1-5 µl	<500 ng
Nuclease-Free Water to		25 µl

## For a 50 μl reaction volume:

	Component	Volumo	Final
component		volume	Concentration
	2X Taq Master Mix	25 µl	1X
	Upstream Primer, 10 µM	1 µl	0.1–1.0 µM
	Downstream Primer, 10 µM	1 µl	0.1–1.0 µM
	DNA Template	1-5 µl	<500 ng
	Nuclease-Free Water to		50 µl

4. Gently mix the reaction and spin down in microcentrifuge.

#### 5. Set up program for a routine PCR reactions:

Initial Denaturation	94-95°C for 1-5 min
25-40 cycles	94-95°C for 30 sec
	45-70°C for 10-30 sec
	72°C for X min(1min/kb)
Final extension	72°C for 7 min
Final soak	4-10°C

- For a simplified hot start, proceed as described in step 7.
  Otherwise, place the PCR tubes in the thermal cycler and start the cycling program.
- Simplified hot start: Start the PCR program. Once the thermal cycler has reached 94°C, place the PCR tubes in the thermal cycler. In many cases, this simplified hot start improves the specificity of the PCR.